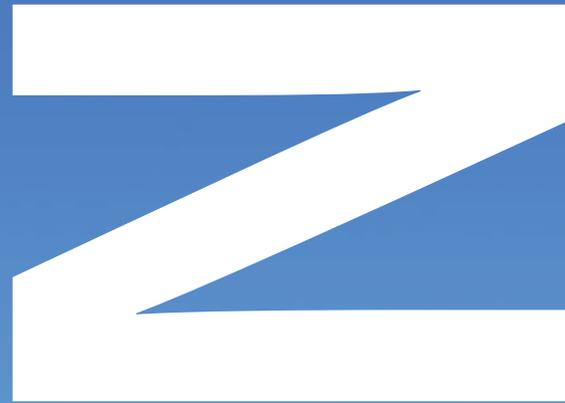
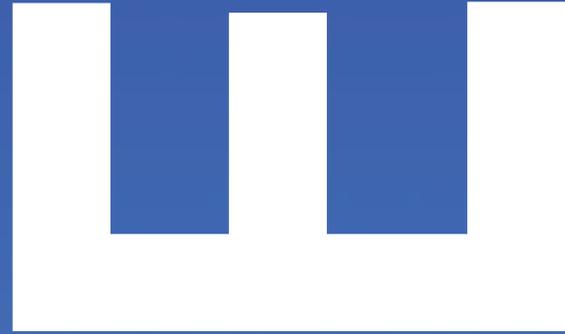




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# Machine Learning-Based Assessment of Invasive Crayfish *Procambarus clarkii* in Riverine Wetlands of the Yeongsangang River Basin, South Korea

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## ABSTRACT

The Invasive red swamp crayfish, *Procambarus clarkii* is rapidly expanding in Korean inland waters, yet its population size and habitat use remain poorly quantified. This study developed a machine learning-based framework to assess the distribution and population size of *P. clarkii* in gravel-bar and abandoned-channel wetlands of southern Korea. Environmental and capture data were compiled for 86 wetlands along the Yeongsangang River and Jiseokcheon Stream surveyed between 2021 and 2023. A Random Forest model with balanced presence-absence sampling and repeated five-fold cross-validation was applied to predict wetland-level habitat suitability. The best model achieved an area under the curve of 0.836 and identified channel and pool density, available habitat area, total wetland area, and permanent surface water bodies as the most important predictors of occurrence. Population density was then estimated from baited umbrella-shaped hexagonal traps with six entrances in eight wetlands, assuming a 4 m effective capture radius and a capture probability of 0.265, and extrapolated to 36 wetlands classified as high or moderate suitability or with confirmed occurrence. Median estimates indicated that approximately  $8.2 \times 10^5$  individuals (range:  $6.1 \times 10^5$ – $1.6 \times 10^6$ ) inhabit these wetlands. Dense populations were associated with large wetlands that maintain permanent surface water, highlighting such sites as priorities for control. Overall, this study demonstrates that integrating habitat suitability modelling with trap-based density estimation can support tiered management of invasive crayfish under limited resources.

**Keywords:** Habitat suitability modeling, Invasive species, Machine learning, Population estimation, *Procambarus clarkii*

## Introduction

Invasive species pose a significant environmental challenge, as they can disrupt the ecological equilibrium of native ecosystems and contribute to biodiversity loss (Gentili *et al.*, 2021). These species exert a deleterious effect on native species through habitat competition, ecosystem disruption, and disease transmission, with some species even dominating ecosystems due to rapid population growth (Gherardi, 2006; Lee & Park, 2019).

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The red swamp crayfish (*Procambarus clarkii*) demonstrates resilience in environments with low oxygen levels and exhibits a notable tolerance to variations in water temperature (Loureiro *et al.*, 2015; U.S. Fish & Wildlife Service, 2015). These characteristics are the reason why *P. clarkii* has experienced such rapid proliferation, while native crayfish population levels have declined due to specific environmental changes (Chucholl, 2013). *P. clarkii* demonstrates a broad spectrum of dietary preferences and behavioral patterns that differ significantly from native crayfish. When competing for food and habitat, *P. clarkii* has been observed to physically push out native crayfish (Pintor *et al.*, 2008). *P. clarkii* is notable for its short generation time and high reproductive rate, reproducing multiple times a year. In contrast, native crayfish species frequently undergo reproductive cycles of only one or two instances per year, a pattern that hinders the preservation of their population sizes (Momot, 1995). Consequently, *P. clarkii* exhibits superior environmental adaptability, competitiveness, and reproductive capacity compared to native crayfish. This dynamic suggests a gradual displacement of the native crayfish population by *P. clarkii* within the local ecosystem.

Furthermore, *P. clarkii* carries *Aphanomyces astaci* and the white spot syndrome virus, which have been known to cause mass mortality among native crayfish populations. In particular, *A. astaci* is an aquatic pathogenic fungus that causes crayfish plague. Several crayfish species native to North America, including *P. clarkii*, have been documented as exhibiting resistance to this fungus (Alderman *et al.*, 1990). However, native crayfish species in Europe and Asia lack resistance to this fungus. Consequently, when infected, mass mortalities and sharp population declines have been reported (Brady *et al.*, 2024; Filipová *et al.*, 2013; Kawai & Kobayashi, 2005; Nakata *et al.*, 2002). Given the shared habitat of *P. clarkii* and native crayfish, the pathogen responsible for the decline in native crayfish populations is, most likely, transmitted through the water. This has led to the hypothesis that *P. clarkii* may become the sole surviving species within the ecosystem (Gherardi, 2006; Holdich *et al.*, 2009).

*P. clarkii* was initially documented in South Korea in 1987 at Yongsan Family Park in Seoul. Subsequently, in 2006, minimal numbers of this species were reported in certain regions (Kim *et al.*, 2006). The official establishment of *P. clarkii* within the domestic ecosystem was confirmed in Jiseokcheon Stream, located in Naju City, Jeollanam-do, in 2019 (Kim *et al.*, 2019). Subsequently, the species has expanded its habitat to include the Yeongsangang, Seomjingang, and Mangyeonggang Rivers. Furthermore, its presence has been substantiated in minor streams near urban areas, including Cheongju City (Lee & Park, 2019; Park *et al.*, 2020). In response, the Ministry of Environment of South Korea classified *P. clarkii* as an

ecosystem-disrupting species in 2019 and is promoting research on population monitoring and preventing its spread.

A quantitative understanding of the distribution and population dynamics of invasive species is essential for effectively controlling it (Ricciardi, 2007; Stohlgren & Schnase, 2006). However, quantitative research on the population of *P. clarkii* in South Korea is lacking. Existing studies have focused on reporting whether it has established itself in natural ecosystems and the number of individuals captured (Choi *et al.*, 2021; Jung *et al.*, 2022; Kim *et al.*, 2019; Park *et al.*, 2020). Population size can be estimated using the capture-recapture method (Efford & Fewster, 2013; Petit & Valiere, 2006). However, given the dispersal of *P. clarkii* along Korean rivers, this method has limitations, as it can only provide information for specific, limited areas. The population size of *P. clarkii* is subject to significant fluctuations. These are influenced by factors such as predation pressure, survival rates of juvenile crayfish, and environmental changes. This underscores the limitations of rudimentary population surveys in formulating long-term control strategies.

A complete census is the most accurate method, but implementing it is challenging due to time, labor, and resource constraints. Furthermore, it poses a risk of habitat destruction and raises ethical concerns (Krebs, 2009; Sutherland, 2006). Therefore, a novel approach is imperative for predicting long-term changes in the *P. clarkii* population and analyzing stable population fluctuation patterns.

In this study, we used the number of individuals captured in a specific area to estimate the total population across the entire river section, rather than surveying the entire area. Specifically, habitat characteristics were established along the entire river section as spatial information, and population size was estimated by comparing these with points where capture surveys were conducted. The population size was estimated by combining information on the number of individuals caught per trap collected during field surveys with information on the potential *P. clarkii* habitats. The aim of this study was to conduct a quantitative assessment of the *P. clarkii* population, thereby contributing to the establishment of effective control strategies.

## Materials and Methods

### Survey area setting

The survey area included the Yeongsangang River and Jiseokcheon Stream in South Korea, encompassing locations where *P. clarkii* was captured during surveys conducted between 2021 and 2023 (Fig. 1). The analysis subjects were 86 wetlands located along the channels of two rivers and their surrounding areas within the study area.



**Table 1.** Environmental information built on wetland data

Class	Variable	Description	Data type
Site information	UID	• Wetland ID	Numeric
Wetland characteristics	type	• Topographical characteristics of the habitat 1: Old riverine wetland, 0: Sandbar wetland	Categorical
	area	• Total area of each wetland (ha)	Numeric
Land-cover	avail_area	• Potential habitat area in each wetland (ha)	Numeric
	lc_bar	• Presence of sandbar 1: Yes, 0: No	Categorical
Hydrological features	lc_bare	• Presence of bare ground 1: Yes, 0: No	Categorical
	open_water	• Presence of open water 1: Yes, 0: No	Categorical
Ecological factors	lc_water	• Presence of perennial water flow 1: Yes, 0: No	Categorical
	channel_pool	• Channels and pools within a wetland 1: Sparse or absent, 2: Scattered, 3: Abundant	Numeric
Crayfish population	food	• Expected abundance of food source 1: Abundant, 0: Low	Categorical
	predation_risk	• Accessibility for terrestrial predators 1: Low, 2: Moderate, 3: High	Categorical
Crayfish population	pop_trap	• Number of red swamp crayfish caught in traps	Numeric
	pop_net	• Number of crayfish caught with a scoop net	Numeric
	trap	• Number of traps	Numeric
	pop_1trap	• Number of crayfish per trap	Numeric
	pop_total	• Total number of crayfish caught	Numeric
	obs	• Red swamp crayfish presence 1: Observed, 0: Not observed	Categorical

Coding: *food* (0=low, 1=abundant); *predation\_risk* (1=low, 2=moderate, 3=high), derived from consistent interpretation of aerial/satellite imagery and land-cover/topographic maps; *predation\_risk* was based on judged proximity to roads and/or mountainous/forested areas. *avail\_area*, area available for crayfish habitation; *lc\_bar*, presence of gravel/sand bar; *lc\_bare*, presence of bare ground; *open\_water*, presence of open water surface; *lc\_water*, presence of permanent water body; *channel\_pool*, density of channels and pools; *predation\_risk*, accessibility of terrestrial predators; *pop\_trap*, number of individuals captured using traps; *pop\_net*, number of individuals captured using a dip net; *trap*, number of traps deployed; *pop\_1trap*, individuals captured per trap; *pop\_total*, total number of individuals captured; *obs*, indicates red swamp crayfish presence.

abundance of food sources (*food*) and the accessibility of terrestrial predators (*predation\_risk*). *P. clarkii* is classified as omnivorous, meaning it consumes a wide variety of plant material, detritus, and animal prey. The diet of the subject consists of aquatic insects, freshwater shrimp, and aquatic plants in substantial quantities. Consequently, the abundance of food sources (*food*) and the accessibility of terrestrial predators (*predation\_risk*) that constrain them were incorporated as significant ecological factors. The terrestrial predators under consideration included humans, otters, and leopard cats. The proximity to mountainous

regions or roads was determined to be indicative of favorable accessibility. Food availability (*food*) was coded as a binary variable (0/1) to represent relative resource abundance within each wetland. Based on consistent visual interpretation of aerial/satellite imagery and land-cover/topographic maps, wetlands showing clear indicators of abundant food resources (e.g., well-developed aquatic vegetation and/or visible organic detritus/accumulated organic matter) were coded as *food*=1, whereas wetlands lacking such indicators were coded as *food*=0. Predation risk (*predation\_risk*) was coded as a categorical variable

(2/3) to reflect the accessibility of terrestrial predators (e.g., humans, Eurasian otter, and leopard cat). Wetlands were classified as *predation\_risk*=3 (high) when roads and/or mountainous/forested areas were judged to be adjacent to or in close proximity to the wetland, indicating high accessibility of terrestrial predators, and as *predation\_risk*=2 (low) when such features were judged to be distant or not readily accessible. This 2-3 coding was retained to distinguish *predation\_risk* from other binary (0/1) variables, and the same interpretation criteria were applied consistently across all wetlands. The crayfish population data were subsequently organized into the following categories: the number of individuals captured using traps (*pop\_trap*), the number captured using nets (*pop\_net*), the number of traps set (*trap*), the number of individuals captured per trap (*pop\_1trap*), and the total number of individuals captured (*pop\_total*). The environment variables constructed in this manner were organized into categorical or numerical data formats. The environmental information for each wetland constructed in this study is presented in the table in Appendix 1.

These variables were selected based on the biological and ecological characteristics of *P. clarkii*, which inhabits environments characterized by stagnant or slow-flowing currents, such as slow-moving streams, wetlands, and irrigation ditches, and burrows into the substrate for shelter in dry or high-temperature conditions. Therefore, topographical and hydrological variables, including wetland type, area, habitable space area, distribution of gravel-and-sand bars and bare lands, open water surfaces, and water body morphology, are essential for describing crayfish habitat conditions. Furthermore, given the omnivorous nature of *P. clarkii* and its reliance on a diverse array of food sources, including aquatic insects, plants, and other organisms, the abundance of these resources was identified as a pivotal ecological factor. Concurrently, the accessibility of terrestrial predators such as humans, otters, and leopard cats was identified as a pivotal factor influencing population viability. This factor was duly reflected in the study's findings. Meanwhile, *P. clarkii* has demonstrated remarkable environmental resilience, with high tolerance for a broad range of conditions, including water temperatures ranging from 0°C to 40°C and pH levels from 5.8 to 10.0. Consequently, water quality parameters were not incorporated as variables in this study.

The investigator constructed all variables through meticulous observation of various materials, including aerial photographs, land cover and topographic maps, and other pertinent documentation. In particular, the “area available for habitation (*avail\_area*)” was constructed through vectorization. The “*avail\_area*” refers to the area within each wetland that meets the environmental conditions necessary for the red swamp crayfish to inhabit. We delineated the primary habitat of *P. clarkii* as sections characterized

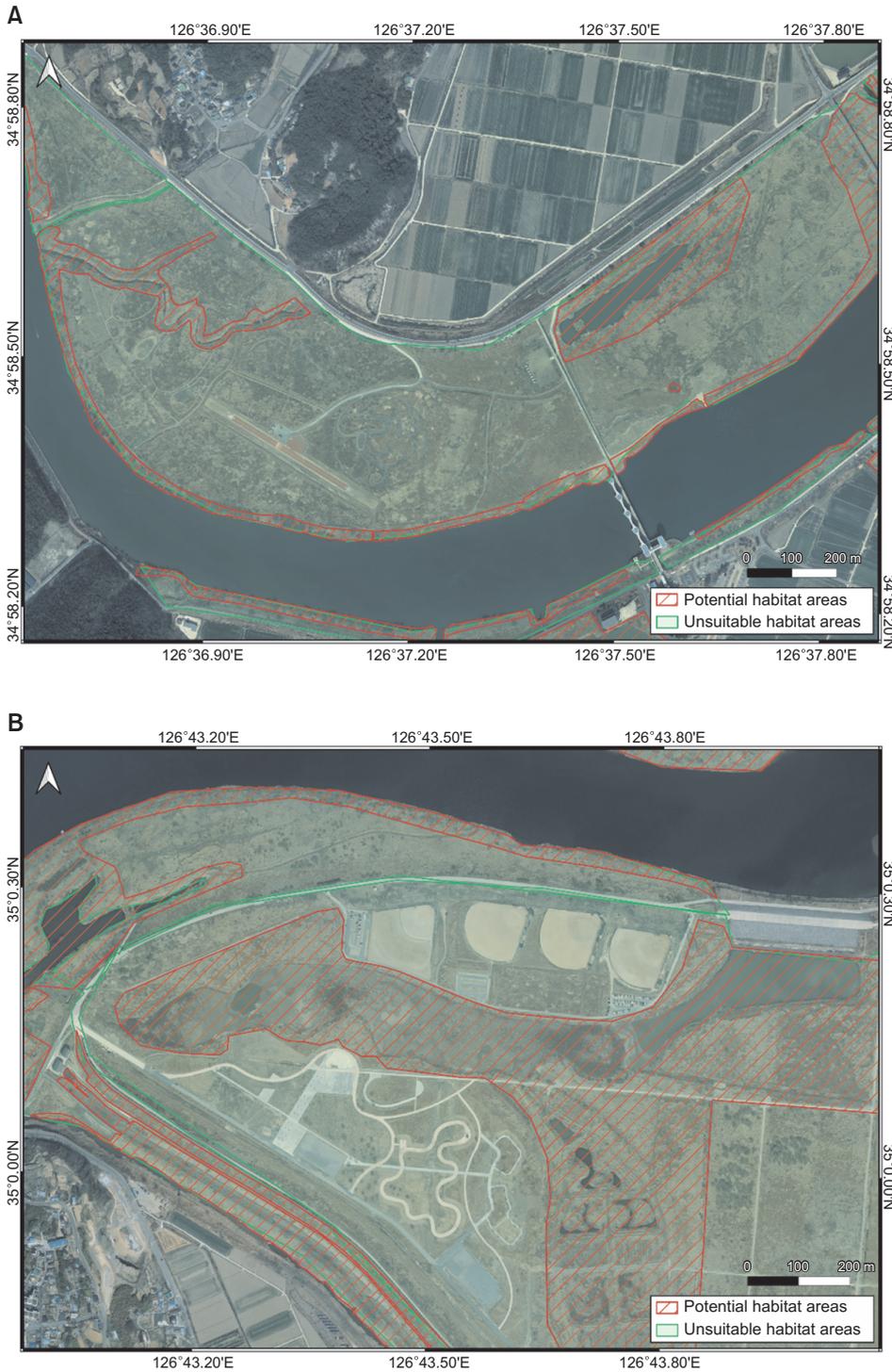
by slow-moving currents and relatively shallow depths (approximately 40-100 cm), based on a literature review (Park *et al.*, 2020). Accordingly, shallow-water areas surrounding gravel-and-sand bar wetlands were designated as potential habitats. In addition, areas meeting the following conditions were excluded from the “area available for habitation.” Areas where the ground surface is covered with concrete or paving materials due to frequent human access or structures such as walkways, roads, bridges, buildings, parking lots, or construction sites (Fig. 2A) and areas where the soil is heavily compacted were excluded because they were deemed unsuitable for the burrowing and habitation of *P. clarkii* (Fig. 2B). The process of classifying wetlands in the study area into “area available for habitation” and “unsuitable areas” was performed by vectorizing the Vworld satellite imagery.

### Study methods

We estimated the *P. clarkii* population size using two steps. First, the environmental data from the 16 wetlands where *P. clarkii* were captured and those from other wetlands were used to assess the probability of *P. clarkii* occurrence using machine learning techniques. Second, the population size within the area available for habitation within wetlands was estimated based on the number of traps and the number of individuals captured across eight wetlands.

In the first step, we used a Random Forest (RF) algorithm to estimate the probability of *P. clarkii* occurrence in wetlands (Fig. 3). Among the 86 wetlands, all 19 wetlands with capture records were included as presence (positive) sites, and 19 wetlands were randomly selected from the 67 wetlands without capture records as absence (negative) sites, yielding a total of 38 wetlands for model development. This subsampling of absence sites was a strategic choice to mitigate the severe imbalance between presence and absence data and to enhance model robustness. When all absence sites were used, the model was disproportionately exposed to negative cases, which hindered learning of the occurrence characteristics of *P. clarkii* and resulted in a strong bias toward predicting absence, ultimately lowering model performance. Accordingly, we constructed a more balanced dataset by limiting the number of absence sites. Model performance was evaluated using the area under the receiver operating characteristic curve area under the curve (AUC).

The selected dataset was partitioned into five folds, and the model was trained and validated using a 5-fold cross-validation framework. Model training was conducted iteratively, and performance was assessed after at least five training runs. A total of 100 repetitions was performed; if the AUC did not meet the threshold of 0.7, model parameters were adjusted and the training procedure was repeated. Repeating cross-validation and prediction 100

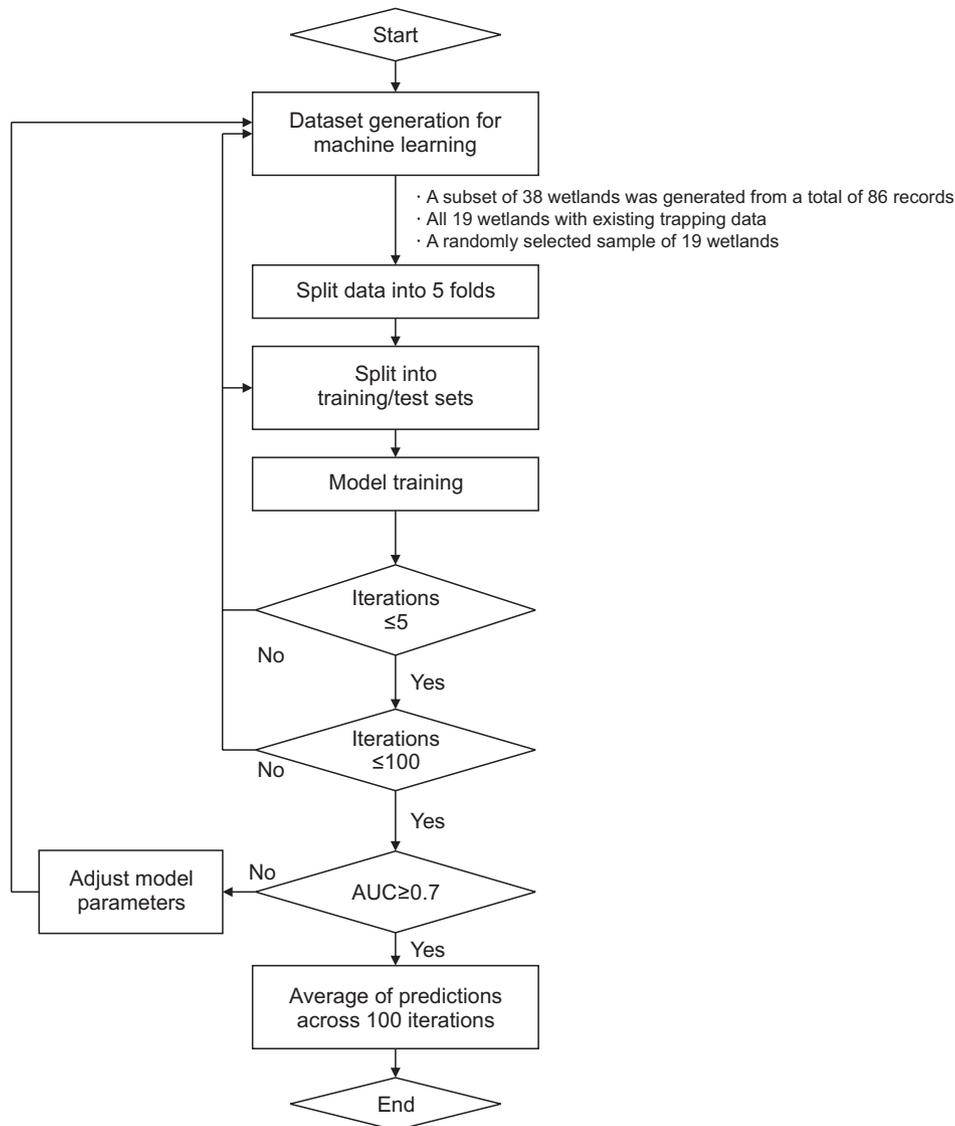


**Fig. 2.** Examples of unsuitable habitat areas. (A) Surfaces covered with concrete or pavement. (B) Compacted soil areas unsuitable for crayfish habitat.

times was intended to minimize stochastic variation arising from random data partitioning and model instability and thereby improve the reliability of the results. Because model performance can vary depending on random train/test splits and hyperparameter initialization even with the same dataset, we averaged predictions across repeated runs to obtain more stable and generalizable estimates.

Specifically, for each repetition, predicted probabilities were generated for the validation (out-of-fold) data; these out-of-fold predicted probabilities were accumulated for each wetland and then averaged to produce the final occurrence probability.

In the second step, population size in wetlands with potential *P. clarkii* occurrence was estimated by con-



**Fig. 3.** Population estimation process using the Random Forest model. AUC, area under the curve.

verting trap catches to areal density using an effective sampling-area approach. Trap catches were obtained using the umbrella-shaped hexagonal automatic traps with six entrances described in Section 2.2; however, the catch was assumed to represent individuals originating from within a surrounding capture radius ( $r$ ), such that the effective sampling area was  $\pi r^2$ . Following Acosta and Perry (2000), we set  $r=4$  m and the capture probability  $P=0.265$ , and estimated density (individuals  $m^{-2}$ ) from the mean number of individuals captured per trap ( $\bar{c}$ ) as  $D \approx \bar{c} / (p \times \pi r^2)$  (Eq. 2). Total abundance for each wetland was then calculated by multiplying  $D$  by the area available for habitation ( $a$ ):  $N_{\text{total}} = a \times D$  (Eq. 3). The population size in the eight wetlands with *P. clarkii* capture data was calculated by multiplying the number of individuals per trap by the area with potential habitat. For the remaining wetlands lacking *P. clarkii* capture data, population size

was calculated as a range using the median, 25th percentile, and 75th percentile values of density (individuals  $m^{-2}$ ) converted from trap catches (Eq. 2) (Fig. 4).

The sphere of influence of the trap had to be assumed to estimate the population size in the surrounding area based on the number captured in the trap. On average, *P. clarkii* moves at approximately 33 cm/min, enabling it to traverse several hundred meters within 24 hours. While *P. clarkii* can travel distances exceeding several kilometers, it is reasonable to consider that in everyday activities, mobility ranges from tens to hundreds of meters. However, it appeared to be an unreasonable assumption that the individual captured in the installed trap traveled approximately several hundred meters (for instance, 400 meters) to reach that location. It is more logical to hypothesize that the individual was captured within a collection radius of several to tens of meters. Therefore, the number

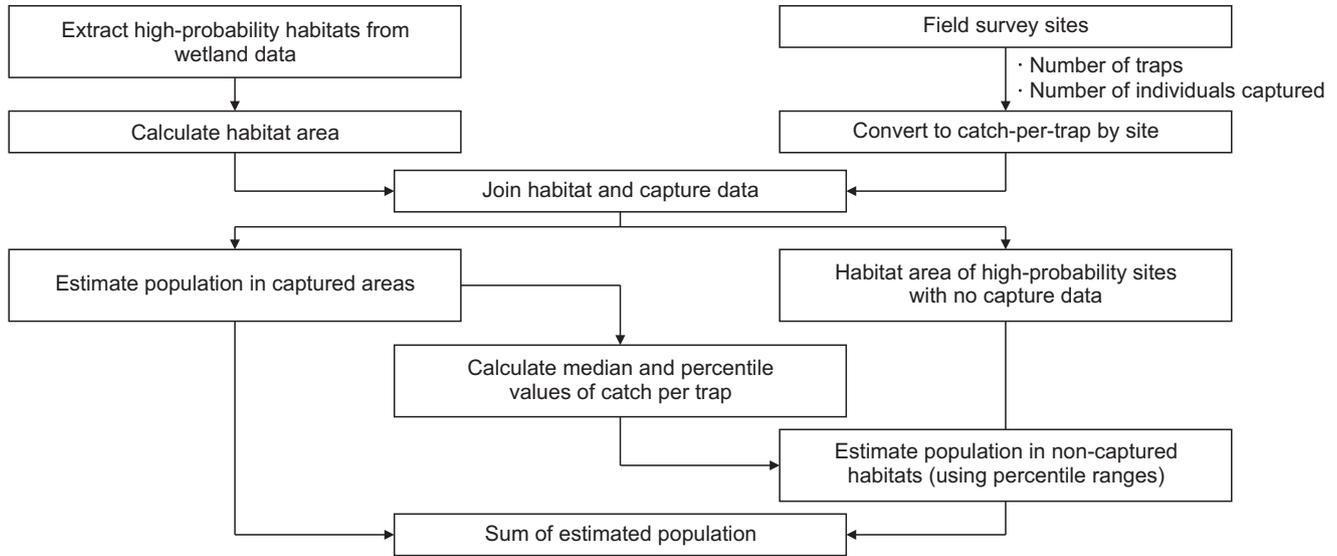


Fig. 4. Flowchart for estimating crayfish population based on habitat suitability and field survey data.

of individuals trapped was estimated using the following equation (Eq. 3).

$$E[c] = p \times N_{local} \quad (\text{Eq. 1})$$

$$D = N_{local} / \pi r^2 = E[c] / (p \times \pi r^2) \approx \bar{c} / (p \times \pi r^2) \quad (\text{Eq. 2})$$

$$N_{total} = a \times D = (a \times \bar{c}) / (p \times \pi r^2) \quad (\text{Eq. 3})$$

$E[c]$  is the expected average number of individuals captured. This number can be expressed as the product of the total number of individuals  $N_{local}$  actually present within the capture radius of the trap and the probability of capturing one individual (Eq. 1). Expressing the area ( $m^2$ ) of the trap's capture radius ( $r$ ) as  $\pi r^2$ , the density per unit area  $D$  could be calculated as shown in Eq. 2. The analysis used the mean of the actual number of individuals captured  $\bar{c}$ , rather than the expected number of individuals captured  $E[c]$ . Therefore, if the area ( $m^2$ ) of the potential habitat was  $a$ , the total population estimate  $N_{total}$  was the product of the density  $D$  per unit area and the area  $a$ , as shown in Eq. 3.

In the analysis, the trap's capture radius ( $r$ ) and capture probability ( $p$ ) are important variables for estimating population size. The value of this variable was determined based on the findings of a study conducted by Acosta and Perry (2000). Through experimental recapture of released Florida crayfish (*P. alleni*) in traps set within a specific radius, it was confirmed that the probability of recapture decreased significantly with distance from the release point. Furthermore, the effective capture radius of a single trap was estimated to be approximately 4 m, and the capture probability based on the capture radius was expressed using the following log-linear model (Eq. 4). This study applied this approach, setting the  $r$ - and  $P$ -

values to 4 m and 0.265, respectively. This implied that the number of individuals captured in the study area corresponded to approximately 25.6% of *P. clarkii* present within a 4 m radius (area of approximately 50  $m^2$ ).

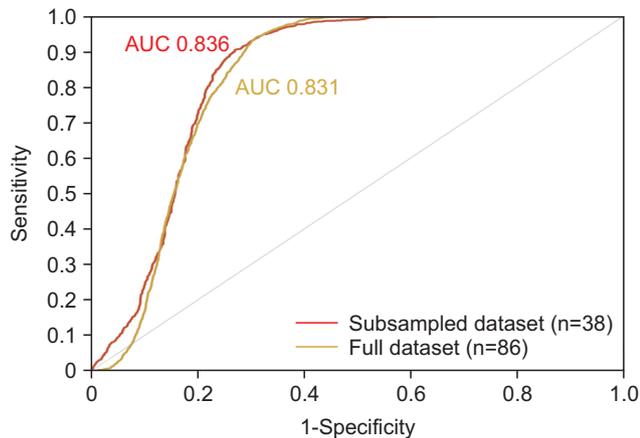
$$P(r) = 0.57 - 0.22 \ln(r) \quad (\text{Eq. 4})$$

## Results

### Estimation of potential wetland habitats

Through 100 iterations of the RF model and 5-fold cross-validation, the effectiveness of the sampling strategy for negative data during model implementation was confirmed via the receiver operating characteristic curve. A random subsampling of 19 negative points balanced against 19 positive points yielded an AUC of 0.836 in the experiment (subsampling dataset). In contrast, results obtained using all negative point data and simply altering the composition of the training and data each time it was repeated yielded an AUC of 0.831 (full dataset). The difference between the two results was minimal (0.005). However, this indicated that the sampling strategy affected the performance of the model (Fig. 5).

The use of the subsampled dataset demonstrated that the potential habitat suitability for *P. clarkii* tended to be determined by the size of the stagnant water areas within wetlands and the available area for habitation (channel\_pool, avail\_area, area), as well as the water persistence ( $I_{c\_water}$ ). Table 2 presents the results of evaluating the importance of variables using Mean Decrease Gini. The channel\_pool (4.70), avail\_area (2.85), and area (2.68) variables had the highest importance scores. This finding suggested that the area of the wetland and the informa-



**Fig. 5.** Comparison of model performance on different datasets. AUC, area under the curve.

tion extracted by crayfish as area available for habitation played a crucial role in determining the presence of the habitat. *lc\_water* (1.41) also emerged as a significant variable, demonstrating that a consistently water-rich environment was a crucial factor for the habitat of *P. clarkii*. The *predation\_risk* (0.46) and *open\_water* (0.44) variables were less important than the previous three variables, yet they still exhibited a significant effect. However, the *type*, *food*, *lc\_bar*, and *lc\_bare* variables showed low importance.

These results indicated that habitat sustainability and the potential for species establishment could increase with sufficient space and stable water resources. This suggested that despite the high environmental tolerance of *P. clarkii*, the importance of area and water resources as fundamental habitat conditions remained a key limiting factor.

Variables that were initially deemed to have minimal importance (*type*, *food*, *lc\_bar*, and *lc\_bare*) could, in fact, exert a non-negligible influence on the habitat of *P. clarkii*. Conversely, the low importance ratings observed could be attributed to limitations in the variable construction process, simplifications in variable definitions, or constraints in the data resolution. For example, the abundance of food sources (*food*) can vary significantly depending on the timing of the survey or the season. Similarly, the distribution of gravel-and-sand basins and bare lands (*lc\_bare*, *lc\_bar*) can also vary in reliability depending on the classification accuracy of the spatial data. Therefore, while the importance results indicate which variables the model used to assess habitat potential, variables with low values are not necessarily irrelevant for habitat prediction.

Fig. 6 shows the estimated potential habitat probability of *P. clarkii* using an RF model. Habitat probability was categorized into three intervals: low (0.12–0.50), moderate

**Table 2.** Variable importance based on Mean Decrease Gini

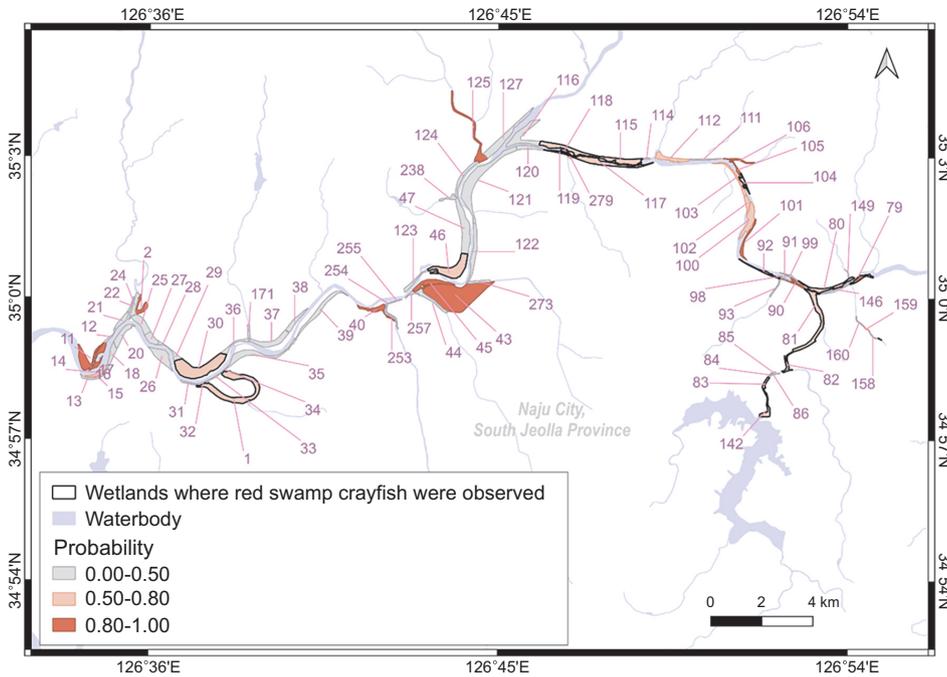
Variable	Gini-based importance
channel_pool	4.70
avail_area	2.85
area	2.68
lc_water	1.41
water_flow	0.44
predation_risk	0.46
open_water	0.44
type	0.13
food	0.04
lc_bar	0.00
lc_bare	0.00

*channel\_pool*, density of channels and pools; *avail\_area*, area available for crayfish habitation; *lc\_water*, presence of permanent water body; *water\_flow*, presence of water flow; *predation\_risk*, accessibility of terrestrial predators; *open\_water*, presence of open water surface; *lc\_bar*, presence of gravel/sand bar; *lc\_bare*, presence of bare ground.

(0.50–0.80), and high-probability (0.80–1.00). Comparing model-based prediction results with actual occurrence points from field surveys revealed that some wetlands were distributed in the high-probability (0.80–1.00) area, consistent with their actual presence, suggesting that the model partially reflects field occurrence characteristics. Conversely, several confirmed wetlands were classified as having a moderate (0.50–0.80) or low probability (0.00–0.50). This was interpreted as arising from variability in field data, uncertainties in the process of establishing environmental variables, and limitations in the training data. Therefore, the map presented in this study should be used as a reference to understand the spatial trends in relative habitat potential rather than as definitive evidence of absolute habitat distribution.

A spatial analysis of wetlands with high potential for *P. clarkii* occurrence revealed that those developed over a larger area on the sedimentary slope exhibit a comparatively higher predicted probability. Abandoned channel wetlands also appeared as areas with high predicted probability. These regional characteristics correspond with the well-established ecological traits of *P. clarkii*, which necessitate stable water resources and sufficient habitat space, while concurrently providing environmental conditions that offer hiding places and diverse food sources. Therefore, these areas may have been designated as habitats with a high probability of the proliferation and dispersal of *P. clarkii*.

However, these predictions inherently contain uncertainty due to the construction process of the environmental variables used, data resolution, and sample size.



**Fig. 6.** Predicted probability of occurrence for *Procambarus clarkii* based on the Random Forest model. High probability wetlands or wetlands whose occurrence has been confirmed were the subject of population estimation.

**Table 3.** Comparison of observed occurrence and predicted habitat suitability probability

Occurrence	Probability			Total
	High	Moderate	Low	
Confirmed	3	13	3	19
Unconfirmed	10	7	50	67
Total	13	20	53	86

For instance, variables such as the abundance of food sources or predation pressure exhibit significant spatio-temporal variability, and their actual influence may have been underestimated during the simplification process. The calculation of variables with low importance does not necessarily imply that they lack ecological significance. This outcome may stem from limitations in the model structure and variable definitions. Therefore, using this map as a foundation for determining absolute distribution is not sufficient. However, considering the efficiency of the simulation, it is likely to serve as a reference material indicating the relative trend of potential habitats for *P. clarkii*.

Table 3 shows the relationship between actual occurrence and predicted probability of occurrence. Of the 86 wetlands examined, *P. clarkii* was confirmed in 19. Of those, three were classified as high probability, thirteen as moderate probability, and three as low probability. This suggested that while the model successfully predicted some actual occurrence points as high or moderate probability, there were still occurrence points classified as low

probability. Meanwhile, for the 67 wetlands where occurrence was not confirmed, 10 were classified as high probability, seven as moderate probability, and 50 as low probability.

In this study, we estimated the population size of *P. clarkii* in wetlands classified as high- to moderate-probability habitats or those where their presence had been confirmed in previous surveys. A total of 13 high-probability wetlands were estimated, and *P. clarkii* was actually observed in three. Moderate-probability wetlands comprised 13 confirmed wetlands and 7 wetlands yet to be identified (Table 3). We selected a total of 36 wetlands as population estimation points, comprising 33 wetlands included in two classes and an additional three wetlands predicted as low probability, but where actual observations were made. The estimated target was finalized through a careful consideration of both the probability of occurrence based on the analysis results and whether it was observed during the field survey.

$$\text{Unconfirmed \& Moderate} = 17 + \text{Confirmed } 19 = 36$$

#### Population size estimation

We estimated the population size of *P. clarkii* across eight habitats where information on the number of individuals caught per trap was confirmed. A total of 1,859 individuals were captured across 295 traps during the 26 surveys conducted in the study area. The number of individuals captured per trap was 6.30. The highest number of individuals confirmed in a single trap was approximately 35, recorded on August 1, 2023. The second-high-

est number of individuals captured was 16.40, recorded on June 22, 2023, in the same wetland area (Table 4).

The total population size was estimated at approximately 327,987 individuals distributed across eight habitats, calculated by multiplying the density per unit area described in the research methods by the area available for habitation (Table 5). This suggests the possibility of a high density of *P. clarkii* in the area.

For the remaining 28 wetlands (17 classified as unconfirmed or moderate probability and above, as well as 11 confirmed wetlands lacking trap information), excluding the eight directly estimated habitats, the population size was estimated by applying the median (1.50), 25th percentile (0.85), and 75th percentile (4.02) of density estimates (individuals m<sup>-2</sup>) derived from trap catches using Eq. 2 to the available area of each wetland.

Table 6 indicates that the estimated population size for

the 28 wetlands was approximately 490,500 individuals. When applying the 25th percentile, the estimate was approximately 277,900 individuals, and when applying the 75th percentile, it was approximately 1,314,500 individuals. Together with the previously estimated population size for the eight wetlands (approximately 7,700 individuals), the total population across the 36 wetlands in the study area was estimated to be approximately 818,500 individuals, with a percentile range from approximately 605,900 to 1,642,500 individuals.

## Discussion

*P. clarkii* possesses ecological traits that enable it to adjust its reproductive strategy, spawning period, brood care, and sexual maturity according to environmental conditions, demonstrating a very high capacity for envi-

**Table 4.** Results of a survey of red swamp crayfish populations in eight wetlands

Wetland ID	Date	Population	Number of trap	Individuals per trap
30	2023.06.22	82	5	16.40
	2023.08.01	421	12	35.08
	2023.08.18	71	7	10.14
	2023.09.14	16	10	1.60
	2023.09.14	1	5	0.20
46	2023.06.22	2	5	0.40
92	2023.02.21	0	1	0.00
	2023.03.28	19	10	1.90
	2023.04.12	94	10	9.40
	2023.08.18	1	10	0.10
	2023.05.17	100	15	6.67
	2023.05.31	139	15	9.27
	2023.05.31	172	15	11.47
	2023.06.22	588	64	9.19
98	2021.06.17	1	1	1.00
	2021.07.05	6	1	6.00
	2021.09.06	1	1	1.00
103	2023.08.01	16	10	1.60
111	2022.08.30	1	5	0.20
117	2023.08.01	2	2	1.00
142	2023.05.17	42	10	4.20
	2023.04.12	21	15	1.40
	2023.05.31	2	15	0.13
	2023.05.25	30	15	2.00
	2023.03.28	11	16	0.69
	2023.08.01	20	20	1.00
Summary statistics (total/average)		1,859	295	6.30

**Table 5.** Estimated crayfish population by eight wetland

Wetland ID	Area (ha)	Potential habitat area (ha)	Average individuals per trap	Estimated population
30	91.7	22.0	15.2	251,044
46	57.4	6.2	0.4	1,862
92	6.4	6.2	8.0	37,236
98	0.7	0.7	2.7	1,419
103	10.4	3.3	1.6	3,964
111	4.3	4.3	0.2	646
117	37.3	30.9	1.0	23,198
142	10.8	8.2	1.4	8,618
Total	219.0	81.8	30.5	327,987

ronmental adaptation (Alcorlo *et al.*, 2008; Loureiro *et al.*, 2015; Sommer, 1984). Since 2019, the Jeonbuk Regional Environmental Office has been capturing over 3,000 *P. clarkii* individuals in 2021 and over 10,000 in 2022. However, their population and range appear to be expanding. The complete eradication of invasive species may create ecological niches that favor new invasions by the same or different species. Consequently, population control is regarded as the optimal control strategy for areas invaded by *P. clarkii* until a more profound understanding of interactions within the invaded ecosystem is achieved (Loureiro *et al.*, 2018).

The estimated population size and growth rate are significant indicators for determining which species should be prioritized for control, as they are related to range expansion and the severity of impacts (Parker *et al.*, 1999; Pintor *et al.*, 2009; Ricciardi, 2003). A greater abundance of invasive species implies increased resource use and interactions with native organisms. Monitoring the abundance of invasive species is therefore crucial for predicting impacts and developing control strategies (Ricciardi, 2007; Stohlgren & Schnase, 2006). Within a biogeographic region, population size is positively correlated with range size (Bock & Ricklefs, 1983; Gaston *et al.*, 1997; Hanski, 1982), and the high abundance of invasive species has been identified as a risk factor for range expansion (Gaston *et al.*, 1997).

Here, we propose an efficient control framework for *P. clarkii* under budgetary and personnel constraints by identifying potential habitats based on ecological characteristics and estimating population size using capture information. The RF-based habitat prediction model showed consistent performance (AUC=0.836) in repeated cross-validation, and was coupled with a trap-to-density conversion to estimate total abundance. Our results indicate that approximately 818,500 *P. clarkii* individuals (605,900–1,642,500) inhabit 36 gravel-and-sand bar wetlands and abandoned channel wetlands in the Yeongsang River and Jiseokcheon Stream basins (median-based

estimates).

Habitat size and the presence of permanent surface water emerged as pivotal factors, suggesting that, despite the high resilience of *P. clarkii*, stable habitat conditions remain a key constraint on establishment. Larger wetlands likely provide greater habitat continuity and refugia (e.g., complex channels and pools), which can buffer populations against local disturbances and increase persistence after partial removal. Permanent surface water may also facilitate overwintering and repeated reproduction by reducing mortality during dry periods and maintaining stable aquatic conditions. From a management perspective, control efforts should prioritize large wetlands with permanent surface water, where populations are more likely to persist and serve as sources for recolonization. Practical measures include intensive trapping/removal in high-suitability sites, repeated follow-up removal to prevent rapid rebound, and targeted surveillance (e.g., seasonal capture surveys or eDNA screening) to enable early detection and rapid response.

However, our estimates depend on the assumed capture radius ( $r$ ) and capture probability ( $p$ ), and uncertainty in these parameters may substantially affect inferred population size. Future studies should calibrate  $r$  and  $p$  through field experiments and quantify uncertainty using bootstrapping or sensitivity analyses. Cross-validation frameworks that account for spatial autocorrelation would further improve model generalizability and reliability.

From a control perspective, this study posits a three-stage control strategy predicated on estimated population size and habitat suitability. First, wetlands with high probability of occurrence, confirmed presence, and large habitat areas (Tier 1) require immediate removal and intensive control of the species. Second, wetlands with a high probability of occurrence but not yet confirmed (Tier 2) require early intervention, necessitating precise monitoring and pilot control experiments. Third, for wetlands with a moderate probability of habitation (Tier 3), low-cost monitoring systems should be established

**Table 6.** Estimated population of *Procambarus clarkii* in the entire study area

Wetland ID	Potential habitat area (ha)	Estimated red swamp crayfish population			Probability	
		25th percentile	Median	75th percentile		
2	11.0	7,266	12,823	34,364	High	
11	17.9	11,824	20,866	55,920		
40	9.2	6,077	10,724	28,741		
43	70.2	46,371	81,831	219,307		
45	14.5	9,578	16,902	45,299		
90	5.9	3,897	6,878	18,432		
101	6.7	4,426	7,810	20,931		
105	4.6	3,039	5,362	14,371		
106	6.5	4,294	7,577	20,306		
125	23.2	15,325	27,044	72,478		
79*	7.7	1,651	2,914	7,810		
14	2.5	10,899	19,234	51,547		Moderate
100	16.5	1,321	2,331	6,248		
102	2.0	8,653	15,270	40,925		
112	13.1	396	699	1,874		
158	0.6	462	816	2,187		
159	0.7	330	583	1,562		
279	0.5	5,086	8,976	24,055		
1*	80.5	53,175	93,838	251,485		
80*	15.2	10,040	17,718	47,485		
81*	31.3	20,675	36,486	97,782		
82*	5.6	3,699	6,528	17,495		
83*	4.6	3,039	5,362	14,371		
104*	6.5	4,294	7,577	20,306		
115*	59.0	38,973	68,775	184,318		
118*	2.3	1,519	2,681	7,185		
119*	2.4	1,585	2,798	7,498		
157*	0.1	66	117	312		
Sum	420.8	277,960	490,520	1,314,594		
Population of Table 5			327,987			
Total (①+②)		605,947	818,507	1,642,581		

Individuals per unit area (ind./m<sup>2</sup>): median, 1.50; 25th percentile, 0.85; 75th percentile, 4.02.

\*Red swamp crayfish observed in field survey.

using eDNA analysis and seasonal capture surveys (Lee *et al.*, 2025). For Tier-1 wetlands, the initial suppression target is set at approximately 10–20% annual population removal to impede further expansion. This target may necessitate adjustment in accordance with the prevailing site conditions, reproductive rates, and the probability of reinfestation. Therefore, the control target should be dynamically adjusted contingent on outcomes from future

field-based capture probability and radius values, as well as sensitivity analysis results (Hein *et al.*, 2007; Prior *et al.*, 2018).

In conclusion, this study has provided a transferable framework for the quantitative assessment of invasive species populations by combining machine learning-based habitat suitability analysis with field capture data. The findings of this study will contribute to the understanding

of the distribution and population dynamics of *P. clarkii* and serve as foundational data for establishing control priorities and developing effective control strategies. This approach can be applied not only in the context of South Korea but also to other regions experiencing problems with the spread of invasive crayfish, providing a useful reference case for international research on invasive species control.

### Author Contributions

Conceptualization: YP, JP. Data curation: YP, SJE, Yunju C, SK. Funding acquisition: Youngho C. Writing – original draft: YP, JP, SK. Writing – review & editing: YP, JP, Youngho C.

### Conflict of Interest

Youngho Cho is an editorial board member of the journal, but they were not involved in the review process of this manuscript. Otherwise, there is no conflict of interest to declare.

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**Appendix 1.** Environmental data for each study wetland

UID	Type	Area	lc_bar	lc_bare	open_water	lc_water	channel_pool	food	predation_risk	avail_area	obs
1	1	80.5	0	0	0	1	3	1	3	80.5	1
2	1	12.6	0	0	1	1	3	1	3	11.0	0
11	0	44.4	0	0	1	1	3	1	3	17.9	0
12	0	3.7	0	0	0	0	1	1	3	0.9	0
13	0	4.3	0	0	0	0	1	1	3	1.3	0
14	0	8.1	0	0	0	0	2	1	2	2.5	0
15	0	1.6	0	0	0	0	1	1	3	0.5	0
16	0	1.6	0	0	0	0	1	1	3	0.4	0
17	0	1.3	0	0	0	0	1	1	3	0.2	0
18	0	14.7	0	0	0	0	1	1	3	5.4	0
20	0	20.8	0	0	0	0	1	1	3	3.8	0
21	0	10.6	0	0	0	0	1	1	3	2.3	0
22	0	3.1	0	0	0	1	1	1	3	1.7	0
24	0	12.8	0	0	0	0	1	1	2	3.9	0
25	0	23.8	0	0	0	0	1	1	3	3.2	0
26	0	31.6	0	0	0	0	1	1	3	2.5	0
27	0	14.3	0	0	0	0	1	1	3	2.2	0
28	0	3.1	0	0	0	0	1	1	3	0.6	0
29	0	6.7	0	0	0	0	1	1	3	1.0	0
30	0	91.7	0	0	0	1	2	1	3	22.0	1
31	0	3.1	0	0	0	0	1	1	3	1.5	0
32	0	2.4	0	0	0	1	1	1	2	0.8	0
33	0	1.8	0	0	0	0	1	1	2	0.6	0
34	0	1.0	0	0	0	0	1	1	2	0.4	0
35	0	39.4	0	0	0	0	1	1	2	8.3	0
36	0	10.0	0	0	0	0	1	1	2	2.8	0
37	0	30.0	0	0	0	1	1	1	2	10.6	0
38	0	14.4	0	0	0	0	1	1	2	4.8	0
39	0	28.6	0	0	0	0	1	1	3	20.3	0
40	0	15.9	0	0	0	1	2	1	3	9.2	0
43	0	185.7	0	0	0	1	2	1	3	70.2	0
44	0	14.8	0	0	1	1	1	1	3	12.1	0
45	0	25.7	0	0	0	1	2	1	2	14.5	0
46	0	57.4	0	0	0	1	2	1	3	20.6	1
47	0	56.9	0	0	0	1	1	1	2	22.2	0
79	0	8.1	0	0	0	1	2	1	3	7.7	1
80	0	20.0	0	0	0	1	2	1	2	15.2	1
81	0	43.5	0	0	1	1	3	1	3	31.3	1
82	0	6.7	0	0	1	1	3	1	3	5.6	1
83	0	4.8	0	0	1	1	3	1	2	4.6	1
84	0	0.4	0	0	0	1	1	1	2	0.3	0

## Appendix 1. Continued

UID	Type	Area	lc_bar	lc_bare	open_water	lc_water	channel_pool	food	predation_risk	avail_area	obs
85	0	0.5	0	0	0	0	1	1	3	0.5	0
86	0	0.8	0	0	0	0	1	1	2	0.6	0
90	0	6.1	0	0	0	1	2	1	3	5.9	0
91	0	1.4	0	0	0	1	2	1	2	1.3	0
92	0	6.4	0	0	0	1	2	1	3	6.2	1
93	0	3.3	0	0	1	1	1	1	3	3.3	0
98	0	0.6	0	0	0	0	2	1	2	0.6	0
99	0	2.8	0	0	0	0	1	1	3	2.0	0
100	0	27.4	0	0	0	1	3	1	3	16.5	0
101	0	11.0	0	0	0	1	2	1	3	6.7	0
102	0	7.8	0	0	0	1	2	1	3	2.0	1
103	0	10.4	0	0	0	1	2	1	3	3.3	1
104	0	6.5	0	0	0	1	2	1	2	6.5	1
105	0	4.6	0	0	0	1	2	1	2	4.6	0
106	0	6.5	0	0	1	1	2	1	3	6.5	0
111	0	4.0	0	0	0	0	2	1	2	4.0	1
112	0	19.4	0	0	0	1	3	1	3	13.1	0
114	0	1.1	0	0	0	0	1	1	2	0.3	0
115	0	61.3	0	0	0	1	3	1	2	59.0	1
116	0	43.6	0	0	0	1	1	1	3	30.7	0
117	0	37.3	0	0	0	1	3	1	3	30.9	1
118	0	2.3	0	0	0	1	2	1	2	2.3	1
119	0	4.1	0	0	0	1	2	1	3	2.4	1
120	0	12.3	0	0	0	0	1	1	3	9.1	0
121	0	92.3	0	0	0	1	1	1	3	70.7	0
122	0	28.1	0	0	0	0	1	1	2	15.0	0
123	0	9.6	0	0	0	0	1	1	3	3.1	0
124	0	15.9	0	0	0	0	1	1	2	6.0	0
125	0	31.3	0	0	1	1	3	1	3	23.2	0
127	0	45.1	0	0	0	0	1	1	3	27.0	0
142	0	10.8	0	0	1	1	3	1	3	8.2	1
146	0	5.6	0	0	0	1	1	1	2	5.6	0
149	0	0.8	0	0	1	1	1	0	3	0.5	0
157	0	0.2	0	0	1	1	2	1	3	0.1	1
158	0	1.1	0	0	1	1	2	1	2	0.6	0
159	0	0.8	0	0	1	1	2	1	2	0.7	0
160	0	1.7	0	0	1	1	2	1	2	1.4	0
171	0	4.3	0	0	1	1	1	0	2	3.7	0
238	0	6.0	0	0	1	1	1	1	3	3.6	0
253	0	4.2	0	0	0	1	1	1	2	2.5	0
254	0	1.4	0	0	0	0	1	1	3	0.8	0

Appendix 1. Continued

UID	Type	Area	lc_bar	lc_bare	open_water	lc_water	channel_pool	food	predation_risk	avail_area	obs
255	0	1.1	0	0	0	0	1	1	3	0.9	0
257	0	2.7	0	0	0	1	1	1	3	1.4	0
273	0	6.9	0	0	1	1	1	1	2	4.5	0
279	0	1.5	0	0	0	1	2	1	3	0.5	0

Coding: *food* (0=low, 1=abundant); *predation\_risk* (1=low, 2=moderate, 3=high), derived from consistent interpretation of aerial/satellite imagery and land-cover/topographic maps; *predation\_risk* was based on judged proximity to roads and/or mountainous/forested areas. No wetlands were classified as *predation\_risk*=1 in this study area. *type*, wetland type; *area*, wetland area; *lc\_bar*, presence of gravel/sand bar; *lc\_bare*, presence of bare ground; *open\_water*, presence of open water surface; *lc\_water*, presence of permanent water body; *channel\_pool*, density of channels and pools; *predation\_risk*, accessibility of terrestrial predators; *avail\_area*, area available for crayfish habitation; *obs*, indicates red swamp crayfish presence.



# Discovery of a Monotypic Genus *Dideopsis* (Diptera: Syrphidae) in South Korea

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## ABSTRACT

The genus *Dideopsis* (Diptera: Syrphidae) is a monotypic genus of the family Syrphidae. *Dideopsis aegrota* (Fabricius, 1805) is reported for the first time from South Korea. This species is mainly distributed in subtropical regions and has never been recorded in temperate regions. Digital image and diagnostic description of this species are provided.

**Keywords:** *Dideopsis aegrota*, Monotypic genus, South Korea, Syrphidae

## Introduction

Syrphidae, commonly known as flower flies or hoverflies, are the most abundant family of the insect order Diptera, with around 6,300 known species worldwide in 209 genera (Mohammadi-Khoramabadi *et al.*, 2024; Sai Teja & Ganiger, 2023). Syrphid flies provide crucial ecosystem services as pollinators, biological control agents and in environmental assessment (Kevan, 2002; Mengual & Thompson, 2011; Ssymank *et al.*, 2008).

The genus *Dideopsis* Matsumura, 1917 is a monotypic genus of the tribe Syrphini. Mengual *et al.* (2008) included *Dideopsis aegrota* in their molecular analysis, and *Dideopsis* was resolved as sister group of *Erionza syrphoides* and *Dideoides coquilletti*. This species is easily distinguished by body size and morphological characteristics.

In this paper, *Dideopsis aegrota* (Fabricius, 1805) is reported for the first time from South Korea. Digital image

and diagnostic description of this species are provided.

## Case Report

The morphological terminology and interpretations used in this study largely follow Thompson (1999) and McAlpine (1981). A photograph were obtained using Nikon D7500 digital camera, processed using Leica Application Suite software (LAS EZ 3.4, Leica Biosystems, Wetzlar, Germany).

## Results

Family Syrphidae Latreille, 1802

Subfamily Syrphinae Latreille, 1802

Tribe Syrphini

Genus *Dideopsis* Matsumura in Matsumura and Adachi, 1917: 142. Type-species: *Eristalis aegrotus* Fabricius, 1805, orig. des.

*Aegrotomyia* Frey, 1946: 158. Type-species: *Eristalis aegrotus* Fabricius, 1805, orig. des.

***Dideopsis aegrota* (Fabricius, 1805) (Figs. 1, 2)**

(Korean name: nal-gae-mu-nui-kkot-deung-e)

*Eristalis aegrotus* Fabricius, 1805: 243. Type-locality: India

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- Dideopsis hemipennis* Hull, 1945  
*Asarkina pura* Curran, 1928  
*Syrphus infirmus* Rondani, 1875  
*Didea ellenriederi* Doleschall, 1857  
*Syrphus fascipennis* Macquart, 1834

**Diagnostic description (after Vockeroth, 1969)**

Large species with moderately broad abdomen bearing broad yellow fasciae and with very broad brown vitta across middle of wing.

Head: eye bare. Face very slightly receding below, oral opening about 2.5 times as long as broad. Face pale yellow, white pollinose, with rather broad black median stripe extending to antennae. Antenna with basoflagellomere slender, sub oval, 1.3 times as long as broad and 1.3 times as long as first two segments combined (male) or distinctly swollen, tapering slightly to apex, 1.5 times as long as broad and 1.7 times as long as first two segments combined (female).

Thorax: scutum shining black, the notopleural area and sometimes a narrow postsutural margin distinctly whitish-yellow pollinose. Scutellum yellow. Pleura dark brown to black, sub shining, with posterior part of anepisternum, anterior part of anepimeron, and dorsalpart of katepisternum densely white pollinose, sometimes slightly paler in ground colour. Scutum with anterior collar of long erect pile. Dorsal and ventral katepisternal pile patches widely separated throughout. Anterodorsal corner of katepisternum with a tuft of pile. Metasternum with long black pile posteriorly. Posteromedian apical angle of metacoxa with a strong tuft of pile. Wing with broad dark brown vitta across median third or slightly more and with base of wing dark brown or with basal 2/3 of wing dark brown; wing apex beyond brown vitta clear. Vein R4+5 very broadly and indistinctly dipped into cell R4+5.

Abdomen oval, not excessively broad, flattened above,

with strong margin from near base of tergum 2 to apex of tergum 5. Tergum 2 with a pair of large, oval, sometimes subconfluent, yellow maculae which reach the anterior and lateral margins broadly; terga 3 and 4 with anterior half or slightly less yellow, otherwise black, the yellow fasciae slightly but distinctly sinuous posteriorly. Sterna yellow, 2 to 5 with broad apical or subapical black fascia which extend over the lateral membrane.

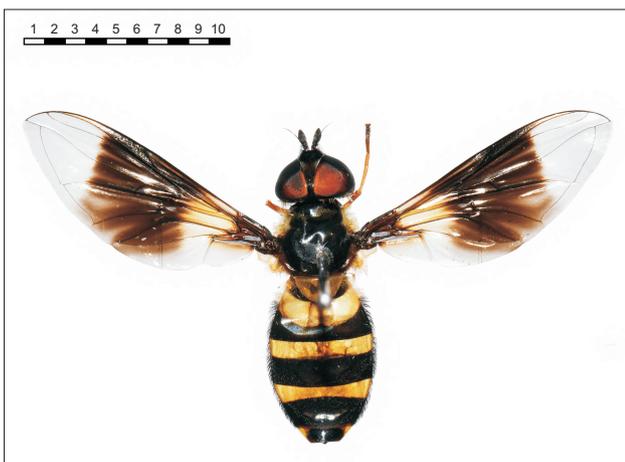
Male genitalia: surstylus broad, short, more strongly broadened and broadly rounded apically, with very abundant setae on inner surface. Sternite 9 slightly narrowed posteriorly, rather strongly produced ventrad below, posteroventrally with a broad deep anteriorly narrowed emargination, laterally with a few short but distinct scattered setae. Superior lobe thick basally and ventrally, semicircular in profile but with posteroventral margin produced dorsad as a strong projecting tooth, ventrolateral margin with a few minute setae. Aedeagal base in the form of a shallow trough with widely divergent, depressed, broadened, dorsal margins and a pair of slender, slightly depressed, subparallel, widely separated, ventrolateral processes. Distal portion of aedeagus. moderately compressed, bent rather strongly ventrad, with depressed sub circular posterodorsal process at the bend, the apex membranous, not inflated, with a few rather stout spicules anteriorly.

**Specimens examined**

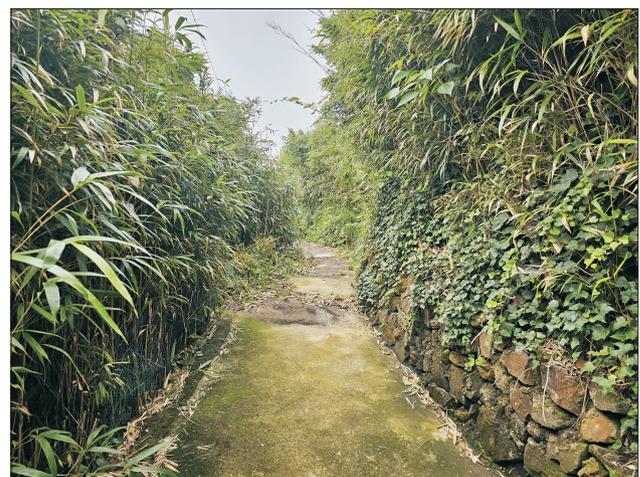
1♀, South Korea: Jeollanam-do, Sinan-gun, Heuksan-myeon, Bi-ri, Jangdo Wetland (34.678686, 125.377948), 7-X-2024, MP.

**Distribution**

Oriental and Palaearctic region - South Korea (new record), China, India, Indonesia, Nepal, Sri Lanka (Ghorpadé, 2014; 2015a; 2015b; Sengupta *et al.*, 2016; Yang *et al.*,



**Fig. 1.** *Dideopsis aegrota*, female body in dorsal view.



**Fig. 2.** Complete view of collected site.

2020).

### Biology

Ghorpadé (1981) recorded this species as a predator of *Aphis spiraecola* (as *A. citricola*), *A. craccivora*, *A. gossypii*, *Pentalonia nigronervosa*, *Rhopalosiphum maidis*, *Toxoptera aurantii*, *T. citricida*, *T. odinae*, aphids on citrus and *Artemisia vulgaris*.

### Discussion

*Dideopsis aegrota* is mainly distributed in subtropical regions and has never been recorded in temperate regions, including Korea. Only one individual of this species was found on an island in southwestern Korea. Therefore, at the present time, it is difficult to determine whether the species is distributed in Korea, flown by air currents, or expanded distribution due to climate change. Therefore, a more comprehensive collection-based study is needed to confirm actual Korean occurrence of this species.

### Author Contributions

Conceptualization: DSC. Data curation: DSC. Formal analysis: DSC, MP. Funding acquisition: DSC, MP. Investigation: MP. Resources: MP. Visualization: MP. Writing – original draft: DSC. Writing – review & editing: DSC, MP.

### Conflict of Interest

The authors declare that they have no competing interests.

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# Import Status of Ornamental Osteoglossiformes in the Republic of Korea

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## ABSTRACT

This study investigates the importation of ornamental Osteoglossiformes into Korea from 2009 to 2024 using National Fishery Products Quality Management Service quarantine records and international databases (CITES, GBIF). A total of 175,797 individuals representing 42 species from six families were imported, with Mormyridae being the most diverse. Imports increased steadily over time, averaging 19.9 species and 6,746.0 individuals annually. Six species, led by *Osteoglossum bicirrhosum* and *Scleropages formosus*, accounted for nearly 80% of total imports. Two species—*S. formosus* (CITES Appendix I) and *Arapaima gigas* (Appendix II)—were identified as internationally threatened but are continuously traded. Imports originated from 16 countries, with Indonesia, Thailand, China, and Nigeria being major suppliers. Ecological risks are highlighted for large carnivorous species such as *A. gigas* and *Chitala ornata*, which have established invasive populations elsewhere, causing declines in native fish. Their potential establishment in Korean rivers, particularly those influenced by warm-water discharge, underscores the need for long-term monitoring and biosecurity measures.

**Keywords:** Biosecurity, CITES, Ecological risk, Import status, Ornamental Osteoglossiformes

## Introduction

Osteoglossiformes, meaning “bony tongue” in Greek, is a relatively primitive order of teleost fishes (Greenwood & Wilson, 1998). All extant species inhabit freshwater bodies across tropical and subtropical regions worldwide (Greenwood & Wilson, 1998). Globally, Osteoglossiformes comprises 241 species in five families, and two species are listed under Cites: *Scleropages formosus* in Appendix I and *Arapaima gigas* in Appendix II (CITES, 2025; Nelson *et al.*, 2016). Most species are large (in body length) and

carnivorous, marking them highly valued in the ornamental fish market (Nelson *et al.*, 2016), and the market for large ornamental fish is rapidly expanding (França *et al.*, 2017; Garcia *et al.*, 2014).

Representative species include *A. gigas*, *Chitala ornate*, *Osteoglossum bicirrhosum*, and *S. formosus*. Species belonging to the genera *Arapaima* and *Chitala* have been reported as invasive in several countries (Camacho & Taniegra, 2021; Miranda-Chumacero *et al.*, 2012). Invasive species may cause population declines or even extinctions of native fauna through predation, competition for resources, habitat alteration (Forneck *et al.*, 2016), or the introduction of novel pathogens (Putra *et al.*, 2018). One of the major pathways for the introduction of non-native species is the global pet trade (Early *et al.*, 2016; Novák *et al.*, 2020). Freshwater organisms kept as pets for ornamental purposes have been known to escape or be intentionally or accidentally released from aquaria (Marková

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et al., 2020).

However, the 2014 non-native species list included only 15 species of Osteoglossiformes, and information on their import status or countries of origin is unavailable (Bang et al., 2014; NIE, 2021). Systematic management and assessment of ornamental fish that may pose potential biosecurity threats are required; therefore, securing baseline data on ornamental fish being imported into Korea is essential.

Accordingly, this study aims to investigate the import records of ornamental Osteoglossiformes into Korea, update the list of non-native fish species, and provide baseline data for future ecological risk assessments and biosecurity management strategies.

### Case Report

To examine the import status of ornamental freshwater aquatic animals we used import/export quarantine statistics from the National Fishery Products Quality Management Service (NFQS, 2025) of Korea from 2009 to 2024 and analyzed the import status of legally confirmed imported ornamental freshwater aquatic animals. The taxonomic classification of imported Osteoglossiformes, and whether they are considered internationally threatened species, were obtained from Global Core Biodata Resource and Convention on International Trade in Endangered Species of Wild Fauna and Flora (CITES, 2025; GBIF, 2025) databases.

### Discussion

#### Status of Osteoglossiformes imports into the Republic of Korea

An analysis of ornamental Osteoglossiformes imported into Korea from 2009 to 2024 revealed that a total of 175,797 individuals representing 42 species across six families have been imported. Among the imported Osteoglossiformes, Mormyridae constituted the largest share

with 25 species (59.5%), followed by Notopteridae (eight species), Osteoglossidae (five species), Arapaimidae (two species), and Gymnarchidae and Pantodontidae (one species each) (Appendix 1). Osteoglossiformes are not recorded in the National Species List of Korea (NIBR, 2025), while the 2014 non-native species list included 15 species: six Notopteridae, five Osteoglossidae, two Arapaimidae, and one Gymnarchidae, and one Pantodontidae (Bang et al., 2014; NIE, 2021).

Examining the data by year, the average number of species imported per year was  $19.9 \pm 2.9$ . Since imports were confirmed in 2009, the number of species has continuously increased, except for a slight decrease between 2018 and 2019 (Fig. 1A). The average number of individuals imported per year was  $6,746.0 \pm 2,400.9$ , and except in 2021, annual imports never exceeded 10,000 individuals (Fig. 1B). Large-bodied species were previously sold only in aquarium shops, but with the growing popularity of online media and e-commerce, their potential market appears to be expanding (Magalhães et al., 2017).

A total of 16 countries exported ornamental Osteoglossiformes to Korea: Indonesia, Congo, Thailand, China, Nigeria, Germany, Singapore, Taiwan, Guinea, Malaysia, Colombia, Peru, India, Japan, Kenya, and Vietnam (Appendix 1). Among these countries, Indonesia accounted for the largest number of species exported (20), followed by Congo (18), Thailand (17), China and Nigeria (15), and Germany (14). Collectively, these top five countries accounted for more than 30% of all imported species (Fig. 2A). By the number of individuals imported, Indonesia led with 64,008 (36.4%), followed by Thailand with 42,236 (24.0%), China with 24,872 (14.1%), and Nigeria with 10,750 (6.1%); collectively, these four countries accounted for 80.6% of all imported individuals (Fig. 2B).

#### Major species

Among imported Osteoglossiformes, *O. bicirrhosum* recorded the highest number of imports (66,921 individuals; 38.1%), followed by *Chitala chitala* (22,763; 12.9%),

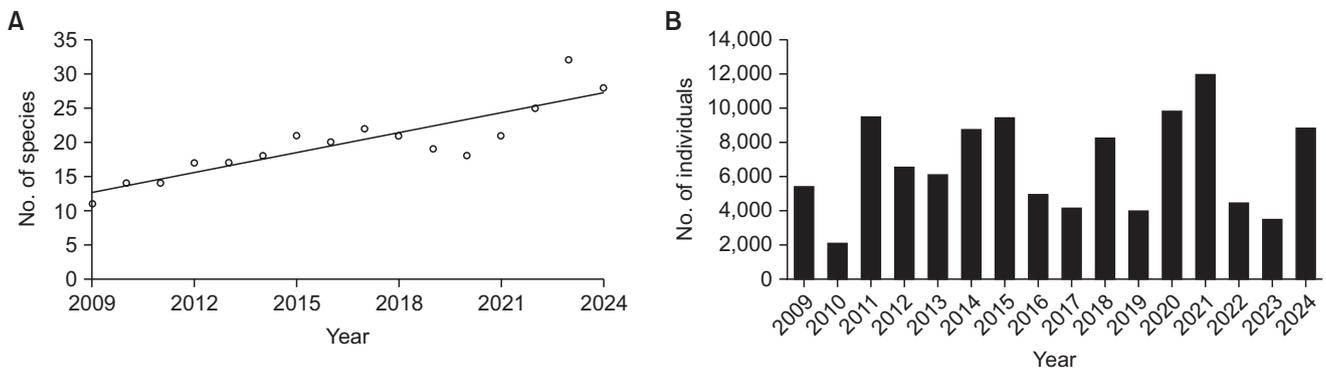
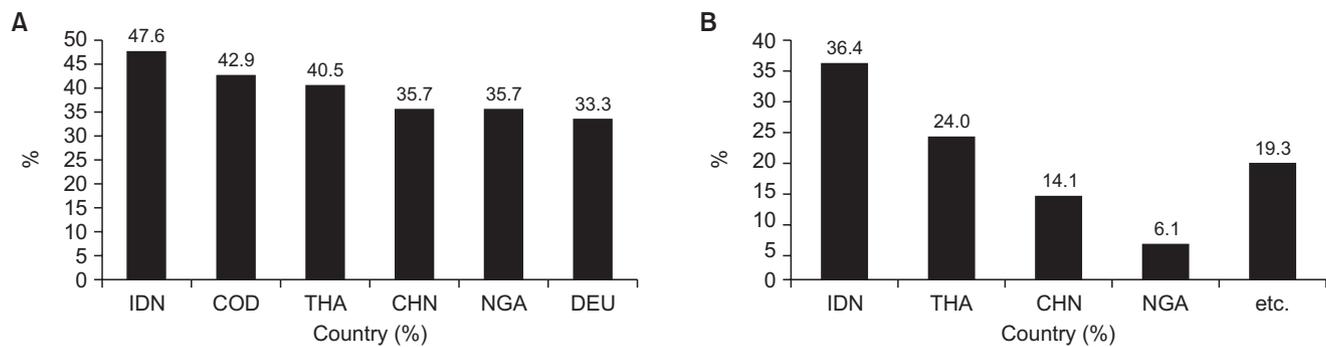


Fig. 1. Annual import status of ornamental Osteoglossiformes. (A) Number of species, (B) number of individuals.



**Fig. 2.** Major importing countries of ornamental Osteoglossiformes. (A) Percentage of species, (B) percentage of individuals. IDN, Indonesia; COD, Democratic Republic of the Congo; THA, Thailand; CHN, China; NGA, Nigeria; DEU, Germany.



**Fig. 3.** Major imported species of ornamental Osteoglossiformes. Photos licensed under CC BY-NC 4.0 via GBIF: (A) *Osteoglossum bicirrhosum* (Tong, 2021); (B) *Scleropages formosus* (Han, 2022); (C) *Chitala chitala* (Leela123\_nunna, 2022); (D) *Chitala ornata* (Wingedg2, 2017).

*Chitala ornata* (15,512; 8.8%), *S. formosus* (13,869; 7.9%), *Gnathonemus petersii* (10,344; 5.9%), and *Mormyrus longirostris* (10,256; 5.8%). These six species together accounted for 79.4% of the total imported individuals (Fig. 3, Appendix 1) (Han, 2022; Leela123\_nunna, 2022; Tong, 2021; Wingedg2, 2017). Two of the imported species (4.7%) were identified as internationally threatened under CITES (Appendix 1). *S. formosus* has been continuously imported since 2009 despite being listed in CITES Appendix I, which includes species threatened with extinction for which international commercial trade is generally prohibited. *A. gigas* has also been continuously imported since 2010 and is listed in Appendix II, which covers species not necessarily currently threatened with extinction but whose trade must be regulated to prevent unsustainable exploitation. Under the CITES permitting system, Appendix I specimens may be traded only under exceptional, non-commercial circumstances and require both export and import permits, whereas Appendix II specimens re-

quire an export permit (or re-export certificate) issued by the exporting country following verification of legal acquisition and a non-detriment finding (CITES, 2025).

In the ornamental trade of large fish species, *O. bicirrhosum* and *S. formosus* are highly popular among collectors, not only for their size (up to 90 cm or more), varied coloration, shiny scales, and other external traits, but also because they are regarded as symbols of luck and wealth, which contributes to their high market value (Yue *et al.*, 2020).

### Ecological risk

Among the Osteoglossiformes, which are mostly carnivorous and large-bodied, the imported species *A. gigas* and *C. ornata* have already been reported as invasive in certain countries (Camacho & Taniegra, 2021; Miranda-Chumacero *et al.*, 2012).

*A. gigas*, known as the pirarucu, is the largest and most iconic freshwater fish of the South American Amazon,

and can reach up to 4 m in length and 200 kg in weight (Castello *et al.*, 2009; Hrbek *et al.*, 2005; U.S. Fish & Wildlife Service, 2022). It has been heavily exploited as a food source over a long period and is currently listed under CITES Appendix II (CITES, 2025; Hrbek *et al.*, 2007). While climate constraints (minimum lethal temperature, 16°C) may limit the establishment and spread of *A. gigas*, its populations have been confirmed in Peru and Bolivia following its introduction to these regions. In northern Bolivia, declines in native fish populations, including many species of high commercial value, have been reported (Van Damme, 2006). *C. ornata* is also native to the Mekong and Chao Phraya river basins in Southeast Asia and is a large predatory species reaching up to 1 m in length (U.S. Fish & Wildlife Service, 2017). In Laguna de Bay, Philippines, a typhoon in 2009 led to the escape of numerous animals from aquaculture facilities. After this event, *C. ornata* rapidly spread and reached approximately 40% of the total catch within two years, causing severe damage to both native and farmed fish species (U.S. Fish & Wildlife Service, 2017).

These large carnivorous species have few natural predators in recipient habitats. By competing with or directly preying on native fish, they can contribute to declines in native fish populations. The primary pathway for the introduction of such non-native species is the international pet trade (Early *et al.*, 2016; Novák *et al.*, 2020), and freshwater animals kept for ornamental purposes have been known to escape or be intentionally/accidentally released from aquaria (Marková *et al.*, 2020). In Korea, the presence of the gastropods *Melanoides tuberculata* and *Mieniplotia scabra*—popular in the ornamental trade as “clean-up” organisms that help control algae and organic matter and capable of parthenogenetic reproduction—has recently been confirmed in the Jukdang Stream (Park *et al.*, 2024; Park *et al.*, 2025). These species have a high potential to establish in rivers and lakes within tropical and subtropical regions; in Korea, they may also establish in rivers sections influenced by warm-water discharge, underscoring the need for long-term monitoring (Park *et al.*, 2024).

### Author Contributions

Conceptualization: YP. Data curation: YP, DC, MJ. Formal analysis: YP. Funding acquisition: YC. Writing – original draft: YP, YC. Writing – review & editing: YP, YC.

### Conflict of Interest

Youngho Cho is an editorial board members of the journal, but was not involved in the review process of this manuscript. Otherwise, the authors declare that they have no competing interests.

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**Appendix 1.** Import status of ornamental Osteoglossiformes list in the Republic of Korea (2009–2024)

Family	Scientific name	2009	2010	2011	2012	2013	2014	2015	2016	2017	2018	2019	2020	2021	2022	2023	2024	Country	CITES		
Arapaimidae	<i>Arapaima gigas</i>	13	141	141	317	1,062	55	129	620	800	200	500	876	485	909	220	600	CHN, COL, THA, PER	II		
	<i>Heterotis niloticus</i>	102	84	40	387	412	320	324	324	499	443	170	457	305	259	72	417	NGA, TWN, SGP, IDN, CHN, KEN, COD, THA			
Gymnarchidae	<i>Gymnarchus niloticus</i>	7	83	43	568	258	271	126	34	111	300	107	146	39	NGA, TWN, DEU, IDN, CHN, THA						
Mormyridae	<i>Brevimyrus niger</i>										10							DEU			
	<i>Brienomyrus brachyistius</i>														11	9	200	NGA, DEU			
	<i>Campylomormyrus alces</i>										48							COD			
	<i>Campylomormyrus elephas</i>	4	70	3	4	8	10											IDN, CHN			
	<i>Campylomormyrus numenius</i>																	COD			
	<i>Campylomormyrus rhyncophorus</i>									117	116	2						33	DEU, IDN, COD		
	<i>Campylomormyrus tamandua</i>									12	10							3	DEU, IDN, COD		
	<i>Cyphomyrus macrops</i>																	10	COD		
	<i>Gnathonemus petersii</i>	344	186	910	1,000	1,180	912	1,091	228	980	660	346	259	471	512	555	710	NGA, DEU, MYS, SGP, IDN, CHN, COD, THA			
	<i>Isichthys henryi</i>																	23	NGA		
	<i>Marcusenius angolensis</i>							50			50							NGA			
	<i>Marcusenius annamariae</i>																	392	GIN, NGA, COD		
	<i>Marcusenius greshoffi</i>																	15	63	GIN, COD	
	<i>Mormyrops anguilloides</i>																	7	CHN, COD		
	<i>Mormyrops boulengeri</i>																	4	1	16	COD

## Appendix 1. Continued

Family	Scientific name	2009	2010	2011	2012	2013	2014	2015	2016	2017	2018	2019	2020	2021	2022	2023	2024	Country	CITES
	<i>Mormyrops mariae</i>															2		COD	
	<i>Mormyrus kannume</i>				5			50	150	100	132	175	175	135	145	50	150	IDN, THA	
	<i>Mormyrus longirostris</i>	10	135	570	115	663	321	508	1,310	849	931	1,157	778	1,208	905			796 NGA, DEU, SGP, IDN, COD, THA	
	<i>Mormyrus rume</i>						10								4			COD	
	<i>Mormyrus tapirus</i>					50	250	378		20	480	744	519	287	180	180	180	IDN	
	<i>Myomyrus macrodon</i>														25	25	25	IDN	
	<i>Petrocephalus christyi</i>														10	29	29	COD	
	<i>Petrocephalus simus</i>													144				NGA	
	<i>Pollimyrus isidori</i>															80	120	GIN, NGA	
	<i>Pollimyrus nigripinnis</i>		55													100		GIN, NGA	
Notopteridae	<i>Chitala blanci</i>	28	20		21	6	30	79	26	29	49	20	17	26	103	18	18	CHN, THA	
	<i>Chitala chitala</i>	1,898	1,900	2,125	2,260	2,165	1,777	1,742	1,754	795	1,078	1,074	1,016	741	1,011	925	502	NGA, DEU, IND, IDN, CHN, THA	
	<i>Chitala lopis</i>					4			118				2	5	5	5	5	IDN, THA	
	<i>Chitala ornata</i>	392	411	274	308	365	758	937	974	890	920	1,105	1,741	1,593	1,630	1,702	1,512	TWN, SGP, IND, IDN, CHN, THA	
	<i>Notopterus notopterus</i>	1,146	194	313	60	280	378	470	449	420	365	180	86	30	50	50	50	DEU, SGP, IND, IDN, CHN, THA	
	<i>Papyrocranus afer</i>			5		10	1	25	70	32	12	56	41	34	49	49	49	GIN, NGA, DEU, COD, THA	
	<i>Papyrocranus congoensis</i>														3			IDN	
	<i>Xenomystus nigri</i>				15	60	5	10	20	10	135	124	265	265	265	265	265	NGA, DEU, IDN, COD, PER	
Osteoglossidae	<i>Osteoglossum bicirrhosum</i>	3,140	3,560	5,288	4,662	3,620	2,845	4,282	4,007	4,055	4,593	4,530	4,766	4,673	4,905	4,071	3,924	TWN, DEU, MYS, IDN, CHN, COL, THA, PER	
	<i>Osteoglossum ferreirai</i>	584	177	81	22	40	377	111	41	227	182	53	4	320	25	18	18	TWN, SGP, JPN, CHN, COL, THA	
	<i>Scleropages formosus</i>	399	670	682	524	966	356	803	985	952	1,221	979	1,108	1,237	894	1,170	923	TWN, DEU, MYS, SGP, IDN, THA	I

## Appendix 1. Continued

Family	Scientific name	2009	2010	2011	2012	2013	2014	2015	2016	2017	2018	2019	2020	2021	2022	2023	2024	Country	CITES
	<i>Scleropages jardinii</i>	142	452		10	252	28					1	300	20			98	MYS, SGP, IDN, CHN, THA	
	<i>Scleropages leichardti</i>				8	58	5	53	40	88								CHN, THA	
Pantodontidae	<i>Pantodon buchholzi</i>	121	155	179	615	616	285	535	170	800	334	237	102	619	187	232	411	SGP, TWN, DEU, VNM, SGP, IDN, COD	

IDN, Indonesia; COD, Democratic Republic of the Congo; THA, Thailand; CHN, China; NGA, Nigeria; DEU, Germany; SGP, Singapore; TWN, Taiwan; GIN, Guinea; MYS, Malaysia; COL, Colombia; PER, Peru; IND, India; JPN, Japan; KEN, Kenya; VNM, Vietnam.

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## AIMS AND SCOPE

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Any experiments involving animals must be demonstrated to be ethically acceptable and where relevant conform to national guidelines for animal usage in research.

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When the research is carried out in areas for which research permits are required (e.g. nature reserves), or when it deals with organisms for which collection or import/export permits are required (e.g. protected species), the authors must clearly detail obtaining these permits in the Acknowledgments section.

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All listed authors must have contributed significantly to the paper, and all authors must be in agreement with the content of the submitted manuscript and must approve of the final version. Please see “Research and Publication Ethics” for detailed information on authorship and author’s responsibilities.

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The journal requires that all authors disclose any potential sources of conflict of interest. Any interest or relationship, financial or otherwise that might be perceived as influencing an author’s objectivity is considered a potential source of conflict of interest. These must be disclosed when directly relevant or directly related to the work that the authors de-

scribe in their manuscript. Potential sources of conflict of interest include, but are not limited to, patent or stock ownership, membership of a company board of directors, membership of an advisory board or committee for a company, and consultancy for or receipt of speaker's fees from a company. The existence of a conflict of interest does not preclude publication. It is the responsibility of the corresponding author to review this policy with all authors and collectively to disclose with the submission ALL pertinent commercial and other relationships. If there are no competing interests, the statement should read, "The authors declare that they have no competing interests." Additionally, potential conflicts of interest of editorial board members should also be disclosed in the manuscript.

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## MANUSCRIPT SUBMISSION

### 1. Select Article Type

- Original (Research) Articles

Original (Research) Article is publishing research on basic ecological research on terrestrial and aquatic populations, communities, ecosystems, and landscapes, as well as ecological research in general. Manuscripts include a title page (with a running title), abstract and keywords, text (introduction, materials and methods, results, and discussion), author contributions, conflicts of interest, acknowledgments, funding, references, tables, figures, and figure legends.

- Review Articles

Review Article is expected to be focused discussion of defined topics relevant to the scope of PNIE. General remarks are same with original article. Manuscripts include a title page (with a running title), abstract and keywords, text, author contributions, conflicts of interest, acknowledgments, funding, references, tables, figures, and figure legends.

- Case Reports

Unique cases that make an important teaching point or scientific observation may be suitable for case report. This

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Thank you for your interest in PNIE. Note that submission implies that the content has not been published or submitted for publication elsewhere except as a brief abstract in the proceedings of a scientific meeting or symposium.

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The covering letter and the manuscript. The covering letter should be uploaded as a file not for review. Figures and tables should also be uploaded as separate files. Figures and tables should also be uploaded as separate files.

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The top, bottom and side margins should be at least 30 mm. All pages should be numbered consecutively in the top right-hand corner, beginning with the title page.

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- E-mail: [pnie@nie.re.kr](mailto:pnie@nie.re.kr)
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We look forward to your submission.

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Papers are accepted for publication in the journal on the understanding that the content has not been published or submitted for publication elsewhere. This must be stated in the covering letter.

The covering letter must also contain an acknowledgment that all authors have contributed significantly, and that all authors are in agreement with the content of the manuscript. The role of each author should be written.

Authors must declare any financial support or relationships that may pose conflict of interest.

If tables or figures have been reproduced from another source, a letter from the copyright holder (usually the Publisher), stating authorization to reproduce the material, must be attached to the covering letter.

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**Spelling.** The journal uses US spelling and authors should therefore follow the latest edition of the Merriam-Webster's Collegiate Dictionary.

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**Nucleotide sequence data** can be submitted in electronic form to any of the three major collaborative databases: DDBJ, EMBL or GenBank. It is only necessary to submit to one database as data are exchanged between DDBJ, EMBL

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Addresses are as follows:

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### 1. Word Length

The length of an article (including references, tables and appendices) should not exceed 20 printed pages for research papers and invited reviews.

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Divide your article into clearly defined sections. Each subsection is given a brief heading. Each heading should appear on a separate line.

#### 1) Title Page

The title page should contain:

- (i) a short informative title that contains the major key words. The title should not contain abbreviations;
- (ii) a short running title of less than 40 characters;
- (iii) the full names of the authors and ORCID IDs of the authors;
- (iv) the author's institutional affiliations at which the work was carried out.

The present address of any author, if different from that where the work was carried out, should be supplied in a footnote.

#### 2) Abstract and Keywords

All articles must have a brief abstract that states in 250 words or fewer the purpose, basic procedures, main findings and principal conclusions of the study. The abstract should not contain abbreviations or references.

Up to six key words (for the purposes of indexing) should be supplied below the abstract in alphabetical order. For the selection of keywords, refer Medical Subject Heading in Index

Medicus or in internet site, <https://www.nlm.nih.gov/mesh/MBrowser.html>.

### 3) Text

Authors should use the following subheadings to divide the sections of their manuscript: Introduction, Materials and Methods, Results, and Discussion.

#### (1) Introduction

Study rationale and relevant background information should be described clearly and concisely.

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Study materials and methods should be described in the following order: study design, materials and methods.

Ensure correct use of the terms sex (when reporting biological factors) and gender (identity, psychosocial or cultural factors), and, unless inappropriate, report the sex and/or gender of study participants, the sex of animals or cells, and describe the methods used to determine sex and gender. If the study was done involving an exclusive population, for example in only one sex, authors should justify why, except in obvious cases (e.g., prostate cancer). Authors should define how they determined race or ethnicity and justify their relevance.

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#### (3) Results

Study results should be reported in a clear and logical manner.

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The results must be explained in relation to the hypotheses proposed in the Introduction. Keep in mind that the Discussion must not be a mere restatement of the results. Authors must emphasize new and important discoveries of the study and state the conclusions drawn from the results in relation to the purpose of the study. The shortcomings and limitations of the study must also be mentioned.

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Enter all author contributions in the submission system during submission. The contributions of all authors must be described using the CRediT Taxonomy of author roles. Read the policy. To qualify for authorship, all contributors must meet at least one of the seven core contributions (conceptu-

alization, methodology, software, validation, formal analysis, investigation, and data curation), as well as at least one of the writing contributions (original draft preparation, review, and editing). Authors may also satisfy the other remaining contributions; however, these alone will not qualify them for authorship. Contributions will be published with the final article, and they should accurately reflect contributions to the work. The submitting author is responsible for completing this information at submission, and it is expected that all authors will have reviewed, discussed, and agreed to their individual contributions prior to manuscript submission.

• Example of author contributions:

Conceptualization: \*\*\*, \*\*\*. Data curation: \*\*\*, \*\*\*. Formal analysis: \*\*\*, \*\*\*. Funding acquisition: \*\*\*, \*\*\*. Investigation: \*\*\*, \*\*\*. Methodology: \*\*\*, \*\*\*. Project administration: \*\*\*, \*\*\*. Resources: \*\*\*, \*\*\*. Software: \*\*\*, \*\*\*. Supervision: \*\*\*, \*\*\*. Validation: \*\*\*, \*\*\*. Visualization: \*\*\*, \*\*\*. Writing – original draft: \*\*\*, \*\*\*. Writing – review & editing: \*\*\*, \*\*\*.

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The authors must disclose any potential conflicts of interest possibly influencing the research or interpretation of data at the time of submission. In particular, all sources of funding for a study should be explicitly stated. Statements on conflict of interest have no influence on the editorial decision to publish. If there are no competing interests, the statement should read, "The authors declare that they have no competing interests." Additionally, potential conflicts of interest of editorial board members should also be disclosed in the manuscript.

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Any persons that contributed to the study or the manuscript, but not meeting the requirements of an authorship could be placed here. For mentioning any persons or any organizations in this section, there should be a written permission from them.

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#### (1) Journal Article

Sugumaran, M., Saul, S.J., and Ramesh, N. (1985). Endogenous protease inhibitors prevent undesired activation of prophenoloxidase in insect haemolymph. *Biochemical and Biophysical Research Communications*, 132, 1124–1129. [https://doi.org/10.1016/0006-291x\(85\)91923-0](https://doi.org/10.1016/0006-291x(85)91923-0)

#### (2) Book

Chapman, R.F. (1971). *The Insects Structure and Function*, 3rd ed. Elsevier.

#### (3) Web Sites

Chapman, K., and Brown, M. (2010). The future of digital library in Asia. *Digital Libraries*, 7, 111–119. Retrieved May 5, 2010 from <https://www.diglib.org/publist.htm>.

GBIF. (2024). *Global biodiversity information facility*. Retrieved Dec 9, 2024 from [www.gbif.org](http://www.gbif.org).

#### (4) Chapter in a Book

Driever, M. (1993). Maternal control of anterior development in the *Drosophila* embryo. In M. Bate, and A.M. Aris (Eds.), *The Development of Drosophila Melanogaster* (pp. 387–424). Cold Spring Harbor Laboratory Press.

#### (5) Conference Abstract

Hong, K.D., and Kim, L.P. (1997). *The sources and migratory pathway of locusts in Korea*. Paper presented at The 50th Annual Meeting of The Entomological Society of Korea, Seoul, Korea.

#### References in articles

We recommend the use of a tool such as EndNote or Reference Manager for reference management and formatting. EndNote reference styles can be searched for here: <https://www.endnote.com/support/enstyles.asp>. Reference Manager reference styles can be searched for here: <https://www.refman.com/support/rmstyles.asp>.

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Tables should be self-contained and complement, but not duplicate, information contained in the text. Number tables consecutively in the text in Arabic numerals. Type tables on a separate page with the legend above. Legends should be concise but comprehensive – the table, legend and footnotes must be understandable without reference to the text. Verti-

cal lines should not be used to separate columns. Column headings should be brief, with units of measurement in parentheses; all abbreviations must be defined in footnotes. Footnote symbols: †, ‡, §, ¶, should be used (in that order) and \*, \*\*, \*\*\* should be reserved for *P*-values. Statistical measures such as SD or SEM should be identified in the headings.

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Type figure legends on a separate page. Legends should be concise but comprehensive – the figure and its legend must be understandable without reference to the text. Include definitions of any symbols used and define/explain all abbreviations and units of measurement.

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Magnifications should be indicated using a scale bar on the illustration.

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Appendices will be published after the references. For submission they should be supplied as separate files but referred to in the text.

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Unique cases that make an important teaching point or sci-

entific observation may be suitable for case report. Abstract should be written in English within 150 words in one paragraph. This should consist of title page, abstract, text (including Introduction, Case Report, and Discussion), author contributions, conflicts of interest, acknowledgments, funding, references, tables, figures, and figure legends.

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Review Article is expected to be focused discussion of defined topics relevant to the scope of PNIE. General remarks are same with original article. Manuscripts include a title page (with a running title), abstract and keywords, text, author contributions, conflicts of interest, acknowledgments, funding, references, tables, and figures. Abstract should be written in English within 150 words in one paragraph. The text can be written in free style. The review articles should be submitted in the same way as regular papers and received the same review process.

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Revised July 26, 2022

Revised June 23, 2025

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## 2. Review Process

PNIE is a peer-reviewed (single-blind peer review) journal and the manuscripts are reviewed by two referees with abundant research experience in the relevant fields of the submitted manuscript, and the final editorial decision is made by the Editorial Committee, based on the referees' evaluations. After reviewing, the manuscript is returned to the corresponding author for revision, the revised manuscript must be re-submitted within one month. If it is not submitted within the designated period without any special reasons, such manuscript is deemed to be withdrawn from evaluation voluntarily.

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The author should not use ideas, research or records of others without quoting them. The corresponding author is responsible for all the contents of the manuscript. Authors must deposit data, strains or other materials in scientific collections (e.g., culture collections, herbaria, GenBank, etc.) to make it possible to repeat the experiments and perform future research. Research published in the PNIE must have been conducted in accordance with institutional, national and international guidelines concerning the use of animals in research and/or the sampling of endangered species. For the policies on research and publication ethics that are not stated in these instructions, the Guidelines on Good Publication (<https://publicationethics.org/resources/guidelines>), Publication of Scholarly Work in Medical Journals (<https://www.icmje.org/icmje-recommendations.pdf>) and the Principles of Transparency and Best Practice in Scholarly Publishing (<https://doaj.org/bestpractice>).

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The corresponding author is responsible for disclosing any potential conflicts of interest that may influence the interpretation of the study's data or findings. Potential conflicts include, but are not limited to, financial support or affiliations with pharmaceutical companies, as well as academic or institutional relationships that could affect objectivity.

All sources of funding relevant to the research must be clearly stated. If no competing interests exist, the authors should include the following statement:

“The authors declare that they have no competing interests.” Additionally, any potential conflicts of interest involving members of the editorial board should be disclosed within the manuscript.

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When the journal faces suspected cases of research and publication misconduct such as redundant (duplicate) publication, plagiarism, fraudulent or fabricated data, changes in authorship, an undisclosed conflicts of interest, ethical problems with a submitted manuscript, a reviewer who has appropriated an author's idea or data, complaints against editors, and so on, the resolution process will follow the flowchart provided by the COPE (<https://publicationethics.org/resources/flowcharts>). The discussion and decision on the suspected cases are carried out by the PNIE Editorial Board and Research Ethics Committee of the National Institute of Ecology (<https://www.nie.re.kr/nie/main/contents.do?menuNo=200189>).

## 8. Process for Handling Article Correction, Retraction, and Editorial Expression of Concern

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## **11. Archiving**

It is accessible without barrier from KoreaScience (<https://koreascience.or.kr/>) or National Library of Korea (<https://nl.go.kr>) in the event a journal is no longer published.

Enacted August 5, 2020  
Revised June 23, 2025

## Editor's Guide

Editors have the following responsibilities:

- ◆ To acknowledge receipt of submitted manuscripts within a few days of receipt and to ensure the efficient, fair and timely review process of submitted manuscripts.
- ◆ To ensure that submitted manuscripts are handled in a confidential manner, with no details being disclosed to anyone, with the exception of the referees, without the permission of the author, until a decision has been taken as to whether the manuscript is to be published.
- ◆ To invite reviewers, probably considering the use of an author's suggested referees for his/her submitted article, but to ensure that the suggestions do not lead to a positive bias (e.g. co-authors of previous publications, mentor). The editor maintains the right to use referees of his/her own choice.
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Enacted October 26, 2022  
Revised June 23, 2025

## 1. Name of Journal

The official journal title is *Proceedings of the National Institute of Ecology of the Republic of Korea*. Abbreviated title is PNIE.

## 2. Website

i. The URL address of official journal website is <https://accesson.kr/pnie/>.

### ii. 'Aims and Scope' statement

It aims to promote, but is not limited to, the achievements of basic ecological research conducted at home and abroad. The prospective audience is researchers conducting global collaborative research as well as ecological studies in the Asia-Pacific region. The scope is not only basic ecological research on terrestrial and aquatic populations, communities, ecosystems and landscapes but also applied issues such as data science and climate change based on ecological research.

### iii. Readership

For those who study basic ecological studies of terrestrial and aquatic populations, communities, ecosystems and landscapes, as well as those who study applied ecology such as climate change and data science based on ecological studies. Its readership can be expanded to other positions:

- Researchers can obtain the latest topics in domestic and international ecological research;
- Professors can access and adopt a variety of data for education;
- Students can understand the latest trends in ecological research at home and abroad;
- Policy makers can reflect the results of the papers in international ecological research policies.

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vii. pISSN: 2765-2203, eISSN: 2765-2211

## 3. Publishing Schedule

PNIE is published quarterly in a year (February 1, May 1, August 1, November 1). Supplement issues may be published.

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